



# The Effect of Fisetin Supplementation and High-Intensity Interval Training on Neurogenesis Markers in Aged Alzheimer's Model Mice

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## ARTICLE INFO

*Article type:*  
Research Paper

*Article History:*  
Received: 25 Aug 2024  
Accepted: 29 Oct 2024  
Published: 20 Jan 2025

*Keywords:*  
High-Intensity interval training  
BDNF  
Fn1  
Fisetin  
Alzheimer's disease

## ABSTRACT

**Introduction:** Alzheimer's disease (AD) is associated with a marked reduction in brain-derived neurotrophic factor (BDNF) and fibronectin 1 (Fn1). This study investigates the effects of fisetin supplementation combined with high-intensity interval training (HIIT) on these neurogenesis markers in an aged mouse model of AD.

**Methods:** In this experimental study, 30 aged C57BL/6 mice (weight: 30 g) with AD were randomly assigned to one of the five groups: (1) Control, (2) AD, (3) AD + Fisetin, (4) AD + HIIT, and (5) AD + HIIT + Fisetin. Alzheimer's disease was induced in the AD groups by injecting amyloid-beta ( $A\beta_{1-42}$ ) into the hippocampus. The HIIT protocol consisted of a 10-minute warm-up at 50-55%  $VO_2$  max, followed by seven intervals, each comprising 4 minutes at 80-90%  $VO_2$  max and 3 minutes at 65-75%  $VO_2$  max. Fisetin was administered at 20 mg/kg for eight weeks. Data were analyzed using one-way ANOVA with a significance level of  $P \leq 0.05$ .

**Results:** Significant differences were observed in BDNF, Fn1, and  $A\beta$  gene expression levels across the five groups of aged mice ( $p < 0.001$ ). BDNF and Fn1 expression were significantly reduced in the AD groups compared to the healthy controls ( $p < 0.001$ ). However, their expression levels increased significantly in the AD + Training + Fisetin, AD + Training, and AD + Fisetin groups compared to the AD-only group ( $p < 0.001$ ). The AD + Training + Fisetin group exhibited the highest expression levels, followed by the AD + Training and AD + Fisetin groups ( $p < 0.001$ ).  $A\beta$  expression was significantly reduced in all intervention groups, with the AD + Training + Fisetin group showing the most substantial decrease ( $p < 0.001$ ).

**Conclusion:** Combining HIIT and fisetin supplementation may promote cerebral neurogenesis in AD by reducing  $A\beta$  levels and enhancing BDNF and Fn1 gene expression. Notably, the combined intervention of HIIT and fisetin exhibits a more significant effect than either HIIT or fisetin alone, with HIIT being more effective than fisetin as a standalone treatment. Thus, the combination of HIIT and fisetin appears to be the most effective complementary approach for managing this disease.

► Please cite this paper as:

Jalali Dehkordi A, Azamian Jazi A, Jalali Dehkordi Kh. The Effect of Fisetin Supplementation and High-Intensity Interval Training on Neurogenesis Markers in Aged Alzheimer's Model Mice. *J Nutr Fast Health*. 2024; 13(1): 35-43. DOI: 10.22038/JNFH.2024.82094.1528.

## Introduction

Alzheimer's disease (AD), the most prevalent chronic disease among the elderly, is an irreversible neurodegenerative disorder characterized primarily by memory loss and other cognitive impairments. It is the most common form of dementia (1) and is also recognized as a proteinopathy, a condition where proteins become abnormal (2). The hallmark of AD is the formation of dense amyloid plaques, known as senile plaques, in the brain (3). The primary component of these plaques is beta-

amyloid ( $A\beta$ ).  $A\beta_{1-42}$ , a proteolytic product derived from the amyloid precursor protein (APP) (4), is a soluble peptide in the body that can form dense, neurotoxic aggregates through fibrillization and oligomerization (5). AD is a progressive form of dementia, neuropathologically characterized by a significant reduction in the number of neurons, particularly in the hippocampus, along with the deposition of beta-amyloid in blood vessel walls, the spread of neuritic plaques, and the formation of intracellular fibrillar tangles (6). These

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pathological changes affect the cortex, hippocampus, and medial temporal lobes. Additionally, the disease is associated with mitochondrial dysfunction, oxidative neuronal damage, synaptic loss, neuronal degeneration, and cognitive and perceptual impairment (7). Neurotrophins are a small group of structurally and functionally related growth factors (8). The Brain-Derived Neurotrophic Factor (BDNF) has received considerable attention due to its essential role in synaptic plasticity, memory, and neurogenesis. BDNF is recognized as a key factor positively regulated by exercise (9). Low levels of BDNF are associated with cognitive impairments, depression, neurodegenerative diseases, and increased mortality. BDNF has been shown to mediate the beneficial effects of exercise on synaptic plasticity and cognitive function, potentially through its role in energy metabolism (10). Studies indicate that BDNF mRNA levels significantly increase in rats following voluntary exercise, which correlates with improvements in spatial memory (11). Exercise can enhance the expression of BDNF and CREB genes. BDNF may influence memory by inducing changes in membrane receptor expression and trafficking, as well as activating several pathways (PLC- $\gamma$ , PI3K, ERK) critical for synaptic plasticity. The exercise-induced increase in BDNF levels, along with improved mitochondrial function, can reduce oxidative stress (12). Fibronectin (FN) is derived from various cellular sources, including astrocytes, epithelial cells, fibroblasts, and mesenchymal cells, and plays a role in cell adhesion, proliferation, differentiation, epithelial tissue repair, immune regulation, nerve regeneration, and other physiological activities (13). Fibronectin 1 (*FN1*) is an effective vehicle for delivering nerve growth factors (NGF) and provides an ideal matrix for axonal repair. In animal studies, fibronectin has been used to enhance peripheral nerve repair processes (14). It forms cable-like structures that promote cellular connections and facilitate the migration of cells such as fibroblasts, neurons, macrophages, and Schwann cells, thereby contributing to peripheral nerve repair (15). Results have shown that increasing physical activity and regular exercise, even low-intensity activities such as walking, strongly protect the nervous system and reduce the risk of dementia (16). Regular physical activity plays a positive role in aging-associated brain atrophy reduction

and diseases related to cognitive impairment, thereby decreasing the incidence of Alzheimer's disease (17). Physical exercise, particularly aerobic exercise, positively impacts brain health and cognitive function, reducing the detrimental effects of neurological disorders such as Alzheimer's, Parkinson's, and depression (18). The beneficial effects of HIIT on the brain are especially pronounced in the hippocampus and dentate gyrus regions (19). These effects include increased blood flow and hippocampal volume in humans, morphological changes in dendrites and dendritic spines, and enhanced synaptic plasticity and neurogenesis in animal models (19). Recent findings suggest that combined HIIT training and herbal supplementation can produce synergistic benefits in preventing and mitigating AD (20, 21).

Fisetin (3,3',4',7-Tetrahydroxyflavone) is a natural flavonoid in various fruits and vegetables, including strawberries, apples, persimmons, grapes, onions, and cucumbers. It possesses neurotrophic, anticancer, and anti-inflammatory properties, as well as antioxidant effects against cognitive and neurological disorders, such as Alzheimer's disease (AD) (22). Additionally, fisetin has been reported to play a role in preserving neurological function during aging (23). Generally, changes in neurotrophic levels due to aging, genetic factors, and other variables have been observed in neurodegenerative diseases, contributing to neuronal destruction. Given the neuroprotective and nutritional properties of factors such as BDNF and Fn1, which enhance central nervous system function, including memory and learning, and considering the limited studies on the combined effects of exercise and fisetin supplementation on BDNF and Fn1 in neurodegenerative diseases like Alzheimer's, there is a clear need to investigate these impacts. Thus, the present study aimed to examine the effects of fisetin supplementation and high-intensity interval training on specific neurogenesis markers in the brains of aged Alzheimer's model mice.

## Materials and Methods

In this experimental study, 30 aged female C57BL/6 mice were obtained from the animal breeding center at the Royan Institute, Isfahan, and were randomly assigned to the following groups: (1) healthy control, (2) AD, (3) AD +

Fisetin, (4) AD + HIIT, and (5) AD + HIIT + Fisetin. Initially, the mice were acclimated to the laboratory environment for one week. The sample size was determined using a previous study and G\*Power software. During the study, mice were kept under standard conditions, including a 12-hour light/dark cycle, an ambient temperature of 20 to 22°C, relative humidity of 55%, and free access to food and water. Additionally, the study adhered to the ethical guidelines of the Helsinki Declaration, and the protocol was reviewed and approved by the Ethics Committee of Shahrekord University (approval code: IR.SKU.RERC.1402).

To induce Alzheimer's disease, Aβ1-42 oligomers (Sigma-Aldrich, USA) were prepared. Synthetic Aβ1-42 was first dissolved in cold hexafluoroisopropanol (HFIP) (Sigma-Aldrich, USA) for 20 minutes. The solution was then vortexed for 10 minutes to promote the formation of Aβ1-42 monomers. These monomers were subsequently subjected to vacuum spinning and precipitation and then dissolved in a 10% HFIP solution. The Aβ1-42 solution was incubated with continuous stirring at room temperature for 48 hours, followed by centrifugation at 4°C for 20 minutes. The supernatant was separated and transferred to pre-chilled tubes. After complete evaporation of HFIP, 50 μM Aβ1-42 oligomers were obtained. The oligomer solution was stored at 4°C until use. Female BALB/c mice were anesthetized with an intraperitoneal injection of 0.2% sodium pentobarbital (50 mg/kg) and then positioned in a stereotaxic apparatus (Stoelting, Wood Dale, IL, USA). Aβ1-42 oligomers were injected into the hippocampus's bilateral dentate gyrus (DG) in

C57BL/6 mice. The injection was performed with a concentration of 50 μM Aβ1-42 or 0.9% sterile saline at a volume of 1 μL, administered at a rate of 0.2 μL per minute.

#### High-Intensity Interval Training Protocol

The HIIT program for the rats in the exercise group will be conducted on a specialized animal treadmill (Tajhiz Gostar Iranian, 2016, Tehran, Iran). The protocol consists of a 10-minute warm-up period at 50-55% VO<sub>2</sub>max, followed by seven intervals, each comprising 4 minutes at 80-90% VO<sub>2</sub>max and 3 minutes at 65-75% VO<sub>2</sub>max, and concludes with a 1-minute cool-down period. Notably, no electrical shocks will be used during the exercise program. If needed, the animals will be encouraged to continue the exercise through manual guidance or auditory cues applied to the treadmill cover (24).

#### 3-7- Fisetin Supplement

The fisetin supplement was obtained from Sigma-Aldrich (USA) and administered orally via gavage at a dose of 20 mg/kg for eight weeks (19).

#### Dissection and Sampling

Forty-eight hours after the final exercise session and following a 12-hour fasting period, the mice were anesthetized with a mixture of 50 mg/mL ketamine and 20 mg/mL xylazine. Laboratory specialists initially performed a pain response test to confirm adequate anesthesia. Once full anesthesia was verified, the animals' brains were exposed, and brain tissue was extracted. The tissue was immediately placed in liquid nitrogen for preservation.

**Table 1.** Primer Sequences for Research Variables

Genes	Primer Sequences	Temperature	Size (bp)
B2m	Forward: 5'- ACAGTTCCACCCGCCTCACATT -3' Reverse: 5'-TAGAAAAGACCAGTCCTTGCTGAAG -3'	60	105
FN1	Forward: 5'- CCCTATCTCTGATACCGTTGTC -3' Reverse: 5'- TGCCGCAACTACTGTGATTCCG -3'	60	110
BDNF	Forward: 5'- GGCTGACACTTTTGAGCACGTC -3' Reverse: 5'- CTCCAAGGCACTTGACTGCTG -3'	60	123
AB	Forward: 5'- TCCGTGTGATCTACGAGCGCATC -3' Reverse: 5'GCCAAGACATCGTCGGAGTAGT -3'	59	128

#### Gene Expression Measurement for BDNF, Fn1, and Aβ1

RNA was extracted from hippocampal brain tissue using TRIzol reagent, following the manufacturer's protocol (Yekta Tajhiz Azma). Subsequently, cDNA synthesis was carried out according to the BIOFACT kit protocol. Gene

sequences for BDNF, Fn1, and Aβ1, along with the nucleotide sequences of the forward and reverse primers, were obtained from the NCBI database (Table 1). Primers were designed using Oligo7 and Beacon Designer software. Real-time PCR was performed using SYBR Green kits on the Corbett Rotor-Gene 6000 system, with Beta-2-

microglobulin (B2M) serving as the housekeeping gene. Gene expression was analyzed and evaluated using the  $2^{-\Delta\Delta CT}$  method.

### Data Analysis

The Shapiro-Wilk test was used to assess normality, and Levene's test was applied to evaluate the homogeneity of variances. Data analysis was conducted using one-way ANOVA followed by Tukey's post-hoc test, with a significance level set at  $p \leq 0.05$  for all tests. All statistical analyses were performed using SPSS software, version 23.

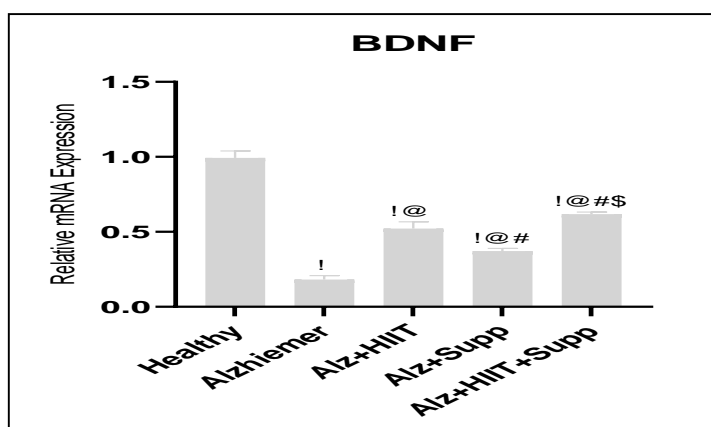
### Results

One-way ANOVA demonstrated significant inter-group differences in BDNF gene expression in the brain tissue of aged mice after eight weeks of HIIT ( $p = 0.001$ ). Tukey's post-hoc test revealed that BDNF expression was significantly lower in the AD group compared to the healthy control group ( $p = 0.001$ ). Additionally, BDNF expression was significantly higher in the AD + Exercise + Fisetin ( $p = 0.001$ ), AD + Exercise ( $p = 0.001$ ), and AD + Fisetin ( $p = 0.001$ ) groups compared to the AD group. Furthermore, BDNF expression was significantly higher in the AD + Exercise + Fisetin group than in the AD + Exercise ( $p = 0.001$ ) and AD + Fisetin ( $p = 0.001$ ) groups. Moreover, BDNF expression in the AD + Exercise group was significantly higher than in the AD + Fisetin group ( $p = 0.001$ ).

Following eight weeks of HIIT, one-way ANOVA indicated significant differences in Fn1 gene

expression levels in the brain tissue of aged mice across the study groups ( $p = 0.001$ ). Tukey's post-hoc test demonstrated a significant reduction in Fn1 gene expression in the AD group compared to the healthy control group ( $p = 0.001$ ). However, Fn1 expression was significantly elevated in the AD + Exercise + Fisetin ( $p = 0.001$ ), AD + Exercise ( $p = 0.001$ ), and AD + Fisetin ( $p = 0.001$ ) groups compared to the AD group. Furthermore, Fn1 expression in the AD + Exercise + Fisetin group was significantly higher than in the AD + Exercise ( $p = 0.001$ ) and AD + Fisetin ( $p = 0.001$ ) groups. Additionally, Fn1 expression in the AD + Exercise group was significantly higher than in the AD + Fisetin group ( $p = 0.001$ ).

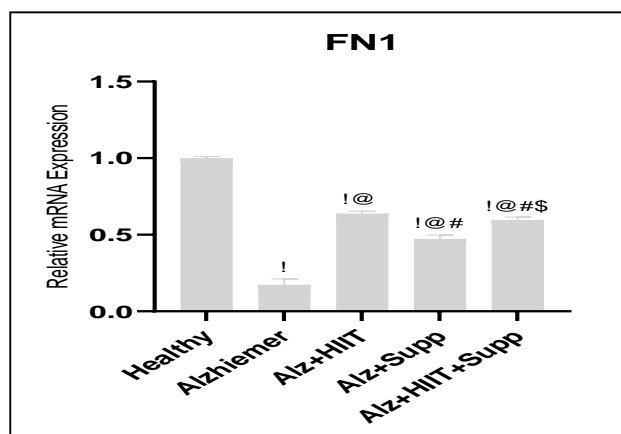
Similarly, after eight weeks of HIIT, one-way ANOVA indicated significant differences in A $\beta$  gene expression levels in the brain tissue of aged mice across the study groups ( $p = 0.001$ ). Tukey's post-hoc test demonstrated a significant increase in A $\beta$  gene expression in the AD group compared to the healthy control group ( $p = 0.001$ ). However, A $\beta$  expression was significantly reduced in the AD + Exercise + Fisetin ( $p = 0.001$ ), AD + Exercise ( $p = 0.001$ ), and AD + Fisetin ( $p = 0.001$ ) groups compared to the AD group. Additionally, A $\beta$  expression in the AD + Exercise + Fisetin group was significantly lower than in the AD + Exercise ( $p = 0.001$ ) and AD + Fisetin ( $p = 0.001$ ) groups. Furthermore, A $\beta$  expression in the AD + Exercise group was significantly lower than in the AD + Fisetin group ( $p = 0.001$ ).



**Figure 1:** Relative expression changes of BDNF gene in different groups

Con: control, AD: Alzheimer's disease, Fis; Fistin, Exe; exercise

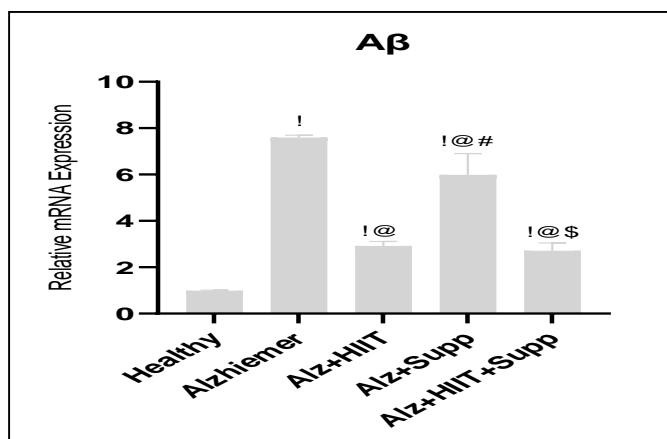
! A significant amount compared to the control group. @A significant amount compared to the Alzheimer's group # Significant level compared to the Alzheimer+HIIT group \$ Significant level compared to the Alzheimer+exercise+physitin group



**Figure 2:** The relative expression changes of Fn1 gene in different groups

Con: control, AD: Alzheimer's disease, Fis; Fistin, Exe; exercise

! A significant amount compared to the Alzheimer's group # Significant level compared to the Alzheimer+HIIT group \$ Significant level compared to the Alzheimer+exercise+physitin group



**Figure 3:** Changes in the relative expression of A $\beta$  gene in different groups

Con: control, AD: Alzheimer's disease, Fis; Fistin, Exe; exercise

! A significant amount compared to the control group. @ A significant amount compared to the Alzheimer's group # Significant level compared to the Alzheimer+HIIT group \$ Significant level compared to the Alzheimer+exercise+physitin group

## Discussion

Aging and AD are associated with reduced brain angiogenesis and neurogenesis. This study examined the effects of HIIT combined with fisetin supplementation on BDNF, Fn1, and A $\beta$  gene expression in elderly mice with AD. In summary, the combination of HIIT and fisetin supplementation, as well as each intervention individually, led to increased expression of BDNF and Fn1 genes and decreased expression of A $\beta$  following eight weeks of HIIT in elderly mice with AD, compared to the Alzheimer's control group. Additionally, the combined effect of fisetin and HIIT was significantly more effective than

either intervention alone, with HIIT demonstrating a more significant impact than fisetin supplementation alone.

Neurotrophic factors, such as BDNF and Fn1, play essential roles in supporting and promoting the growth of various brain neurons. Research indicates that exercise can indirectly influence the expression of neurotrophic factor genes by modulating the release of neurotransmitters such as acetylcholine, gamma-aminobutyric acid (GABA), and monoamines, which in turn leads to elevated BDNF mRNA levels in the hippocampus (25). High levels of BDNF in the hippocampus and cortex reflect its essential role in maintaining proper brain function. BDNF has a clear



neuroprotective role, and exercise enhances cholinergic activity, which is involved in neuronal plasticity induced by physical activity (26). Luo et al. demonstrated that exercise increases neurogenesis in rats and enhances BDNF mRNA production, contributing to improved brain function, learning, and memory (26). Intense physical activity also increases cerebral blood flow, potentially enhancing neuronal cell formation, angiogenesis, synaptogenesis, and neurotransmitter synthesis in various brain regions (27). In a study by Osali et al., BDNF levels and memory improved after 12 weeks of aerobic exercise (28). Zhang et al. (2023) concluded in a review that exercise, mainly treadmill running, swimming, and voluntary wheel running, significantly increases BDNF levels in the hippocampus and cortex of AD models, with swimming being the most effective intervention (29). However, these findings differ from studies by Betio et al. (2020) and Nakhzari et al. (2018), which reported either decreased or unchanged BDNF levels following endurance exercise (30, 31). This discrepancy may stem from differences in subjects or exercise protocols. Another possible mechanism for increased BDNF following HIIT in Alzheimer's mice may involve regulating Trk receptors in brain tissue (32). In the present study, HIIT significantly elevated BDNF levels in the hippocampus of aged Alzheimer's mice.

Fibronectin (FN) is a high-molecular-weight glycoprotein, ranging from 220,000 to 250,000 daltons. As an adhesive molecule, FN was initially isolated from human plasma and fibroblast cell surfaces and is abundantly present in the extracellular matrix (ECM). Fibronectin 1 (FN1) serves as an effective carrier for nerve growth factors (NGFs) and provides a suitable matrix for axonal repair (13). In animal studies, fibronectin has been utilized to enhance peripheral nerve repair processes (33). Research by Ping et al. (2018) highlights FN1's significant role in stimulating the growth, invasion, and survival of gliomas through the activation of the PI3K/AKT signaling pathway (34). Several studies have reported that FN modulates cellular biology via its impact on PI3K/AKT signaling (35, 36). Moreover, research by Lemanska et al. (2009) demonstrated that the molecular status of plasma FN could serve as an additional biomarker for risk assessment (33). Studies show that fibronectin (FN) has a potent

angiogenic effect on endothelial cells in the central nervous system. Additionally, fibronectin enhances the survival and proliferation of brain microvascular endothelial cells, mediated by integrins  $\alpha 5\beta 1$  and  $\alpha V\beta 3$  through mitogen-activated protein kinase signaling (37). The mechanisms underlying the increase in FN1 in the brain after chronic HIIT are likely associated with reduced apoptosis and enhanced PI3K/AKT signaling. This exercise-induced increase in FN1 may also involve the activation of integrins, further supporting the neuroprotective and angiogenic roles of FN1 (33, 34).

The results of the current study indicate that a combination of HIIT and fisetin supplementation, as well as each intervention individually, led to a reduction in A $\beta$  expression after eight weeks of HIIT in aged mice with AD. Mousakhani et al. (2024) demonstrated that voluntary physical activity in an enriched environment likely improves PI3K and Akt protein mechanisms and suppresses amyloid beta accumulation through enhanced insulin sensitivity and hormone function, resulting in cognitive benefits and reduced cell death in AD (37). Additionally, Zarine Afshar et al. (2020) showed that moderate-intensity continuous exercise combined with curcumin supplementation can increase plasma soluble LRP1 levels, promoting A $\beta$  clearance in the hippocampus and modulating factors affecting AD in laboratory rats (38). In contrast, Fallah et al. (2014) reported increased plasma A $\beta$  levels in diabetic rats following voluntary exercise (39), which is inconsistent with our findings. This discrepancy may stem from differences in subjects and exercise protocols. Interval aerobic exercise may reduce brain A $\beta$  plaque levels by regulating amyloid precursor protein processing or enhancing the degradation and clearance of A $\beta$  (37).

In a study, fisetin supplementation significantly improved cognitive performance and memory in mice with AD (22). The current study also demonstrated that fisetin enhances BDNF and Fn1 expression in A $\beta$ 1-42 model mice, thereby supporting critical neurogenic pathways essential for neuronal survival. Notably, fisetin has been shown to reduce apoptosis and neurodegeneration induced by aluminum in the hippocampus of adult mice (40). These results suggest that fisetin plays an essential role in preventing age-related neurodegenerative disorders, including AD (22). A primary

limitation of the present study is the lack of measurement of additional factors related to cognitive performance and brain neurogenesis. Future studies should utilize different measurement techniques, such as Western blotting and ELISA.

## Conclusion

Based on the results, HIIT and fisetin supplementation, either alone or in combination, can enhance neurogenesis in elderly individuals with AD by increasing the expression of BDNF and Fn1 while reducing A $\beta$  levels. Notably, the combination of fisetin and aerobic exercise demonstrates a more substantial effect than either intervention alone, with HIIT alone proving more effective than fisetin alone. Therefore, the combined approach of HIIT and fisetin is recommended as the most effective complementary therapeutic strategy for AD. However, further and more comprehensive studies are required to validate and expand upon these findings.

## Declarations

### Acknowledgements

The authors thank all those who helped in this research.

### Conflict of Interest

The authors declare that no conflicts of interest are associated with this research.

### Funding

Shahrekord University partially supported this research.

### Ethical Considerations

This manuscript is derived from a PhD dissertation in Exercise Physiology at Shahrekord University and has been registered with the Shahrekord University Ethics Committee under the code IR.SKU.RERC.1402.064.

### Author Contributions

All authors equally contributed to the design, execution, analysis of results, and writing of the manuscript.

## References

1. Meamar R, Dehghani L, Ghasemi M, Saadatnia M, Basiri K, Faradonbeh NA, Javanmard SH. Enalapril protects endothelial cells against induced apoptosis in Alzheimer's disease. *Journal of Research in Medical Sciences: the official journal of Isfahan University of Medical Sciences*. 2013;18(Suppl 1):S1.

2. Mayeux R. Epidemiology of neurodegeneration. *Annual Review of Neuroscience*. 2003;26(1):81-104.
3. Hoyer S, Lee SK, Löffler T, Schliebs R. Inhibition of the neuronal insulin receptor An in vivo model for sporadic Alzheimer disease?. *Annals of the New York Academy of Sciences*. 2000;920(1):256-8.
4. Sokoloff L. Relation between physiological function and energy metabolism in the central nervous system. *Journal of Neurochemistry*. 1977;29(1):13-26.
5. Lee MK, Graham SN, Gold PE. Memory enhancement with posttraining intraventricular glucose injections in rats. *Behavioral Neuroscience*. 1988;102(4):591.
6. Ardebili SMM, Yeghaneh T, Ghahesouran J, Rezaazadeh M, Farhoudi M, Ayromlou H, et al. Genetic association of TNF- $\alpha$ -308 G/A and-863 C/A polymorphisms with late onset Alzheimer's disease in Azeri Turk population of Iran. *Journal of Research in Medical Sciences: The Official Journal of Isfahan University of Medical Sciences*. 2011;16(8):1006.
7. Jellinger KA. Recent advances in our understanding of neurodegeneration. *Journal of Neural Transmission*. 2009;116:1111-62.
8. Kuipers SD, Bramham CR. Brain-derived neurotrophic factor mechanisms and function in adult synaptic plasticity: new insights and implications for therapy. *Current Opinion in Drug Discovery and Development*. 2006;9(5):580.
9. Gliwińska A, Czubińska-Łada J, Więckiewicz G, Świętochowska E, Badeński A, Dworak M, et al. The role of brain-derived neurotrophic factor (BDNF) in diagnosis and treatment of epilepsy, depression, schizophrenia, anorexia nervosa and Alzheimer's disease as highly drug-resistant diseases: a narrative review. *Brain Sciences*. 2023;13(2):163.
10. Fakhraei S, Almasi MR, Peeri M, Gharakhanlou R. The effect of 4-Week rehabilitation by aerobic exercise on hippocampus BDNF and TGF- $\beta$ 1 gene expressions in A $\beta$  1-42-induced rat model of Alzheimer's disease. *Journal of Clinical Neuroscience*. 2022;95:106-11.
11. Eggert S, Kins S, Endres K, Brigadski T. Brothers in arms: proBDNF/BDNF and sAPP $\alpha$ /A $\beta$ -signaling and their common interplay with ADAM10, TrkB, p75NTR, sortilin, and sorLA in the progression of Alzheimer's disease. *Biological Chemistry*. 2022;403(1):43-71.
12. King V, Phillips J, Hunt-Grubbe H, Brown R, Priestley J. Characterization of non-neuronal elements within fibronectin mats implanted into the damaged adult rat spinal cord. *Biomaterials*. 2006;27(3):485-96.
13. Kohyama T, Liu X, Wen F-Q, Kobayashi T, Abe S, Ertl R, et al. Nerve growth factor stimulates fibronectin-induced fibroblast migration. *Journal of Laboratory and Clinical Medicine*. 2002;140(5):329-35.
14. King VR, Henseler M, Brown RA, Priestley JV. Mats made from fibronectin support oriented growth of axons in the damaged spinal cord of the adult rat. *Experimental Neurology*. 2003;182(2):383-98.

15. Ten Dijke P, Hill CS. New insights into TGF- $\beta$ -Smad signalling. *Trends in Biochemical Sciences*. 2004;29(5):265-73.
16. Cordingley DM, Marquez I, Buchwald SC, Zeiler FA. Response of Central Nervous System Biomolecules and Systemic Biomarkers to Aerobic Exercise Following Concussion: A Scoping Review of Human and Animal Research. *Neurotrauma Reports*. 2024;5(1):708-20.
17. Debove L, Bru N, Couderc M, Noé F, Paillard T. Physical activity limits the effects of age and Alzheimer's disease on postural control. *Neurophysiologie Clinique/Clinical Neurophysiology*. 2017;47(4):301-4.
18. Abdullahi A, Wong TW-L, Ng SS-M. Understanding the mechanisms of disease modifying effects of aerobic exercise in people with Alzheimer's disease. *Ageing Research Reviews*. 2024:102202.
19. Puoyan-Majd S, Parnow A, Rashno M, Heidarimoghadam R, Komaki A. The Protective Effects of High-Intensity Interval Training Combined with Q10 Supplementation on Learning and Memory Impairments in Male Rats with Amyloid- $\beta$ -Induced Alzheimer's Disease. *Journal of Alzheimer's Disease*. 2024;99(s1):S67-S80.
20. Montero-Odasso M, Zou G, Speechley M, Almeida QJ, Liu-Ambrose T, Middleton LE, et al. Effects of exercise alone or combined with cognitive training and vitamin D supplementation to improve cognition in adults with mild cognitive impairment: a randomized clinical trial. *JAMA Network Open*. 2023;6(7):e2324465-e.
21. Nameni F, Rasoli Z. Investigating the effects of melilot extract and resistance training on protein expression APP in the hippocampus of Alzheimer's model rats. *Sport Physiology & Management Investigations*. 2024;15(4).
22. Ahmad A, Ali T, Park HY, Badshah H, Rehman SU, Kim MO. Neuroprotective effect of fisetin against amyloid-beta-induced cognitive/synaptic dysfunction, neuroinflammation, and neurodegeneration in adult mice. *Molecular Neurobiology*. 2017;54:2269-85.
23. Wang Y, Wu X, Ren W, Liu Y, Dai X, Wang S, et al. Protective effects of fisetin in an A $\beta$ 1-42-induced rat model of Alzheimer's disease. *Folia Neuropathologica*. 2023;61(2):196-208.
24. Gholipour P, Komaki A, Parsa H, Ramezani M. Therapeutic effects of high-intensity interval training exercise alone and its combination with ecdysterone against amyloid beta-induced rat model of Alzheimer's disease: A behavioral, biochemical, and histological study. *Neurochemical Research*. 2022;47(7):2090-108.
25. Maass A, Düzel S, Brigadski T, Goerke M, Becke A, Sobieray U, et al. Relationships of peripheral IGF-1, VEGF and BDNF levels to exercise-related changes in memory, hippocampal perfusion and volumes in older adults. *Neuroimage*. 2016;131:142-54.
26. Gallen CL, D'Esposito M. Brain modularity: a biomarker of intervention-related plasticity. *Trends in Cognitive Sciences*. 2019;23(4):293-304.
27. Lou S-j, Liu J-y, Chang H, Chen P-j. Hippocampal neurogenesis and gene expression depend on exercise intensity in juvenile rats. *Brain Research*. 2008;1210:48-55.
28. Ossali A, Choobineh S, Suri R, Ravasi AA, Mostafavi H. The effect of twelve weeks aerobic exercise with moderate intensity on BDNF, and Short-term memory in 50-65 years old women with syndrome metabolic. *Physiology of Sport and Physical Activity*. 2017;10(1):47-58.
29. Wan C, Shi L, Lai Y, Wu Z, Zou M, Liu Z, et al. Long-term voluntary running improves cognitive ability in developing mice by modulating the cholinergic system, antioxidant ability, and BDNF/PI3K/Akt/CREB pathway. *Neuroscience Letters*. 2024;836:137872.
30. Waller AP, George M, Kalyanasundaram A, Kang C, Periasamy M, Hu K, et al. GLUT12 functions as a basal and insulin-independent glucose transporter in the heart. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*. 2013;1832(1):121-7.
31. Nakhzari Khodakheir J, Haghighi AH, Hamedinia MR. The effects of combined exercise training with aerobic dominant and coenzyme q10 supplementation on serum BDNF and NGF levels in patients with Multiple Sclerosis. *Journal of Arak University of Medical Sciences*. 2018;21(3):94-103.
32. Redolat R, Gresa PM. Reta a tu mente, desafia a tu cerebro: complejidad ambiental y salud cerebral. *Revista INFAD de Psicología International Journal of Developmental and Educational Psychology*. 2016;1(2):201-10.
33. Lemańska-Perek A, Leszek J, Krzyżanowska-Gołąb D, Radzik J, Ka, et al. Molecular status of plasma fibronectin as an additional biomarker for assessment of Alzheimer's dementia risk. *Dementia and Geriatric Cognitive Disorders*. 2009;28(4):338-42.
34. Jing-Ping L. Effects of fibronectin 1 on cell proliferation, senescence and apoptosis of human glioma cells through the PI3K/AKT signaling pathway. *Cellular Physiology and Biochemistry*. 2018;48(3):1382-96.
35. Xiang L, Xie G, Ou J, Wei X, Pan F, Liang H. The extra domain A of fibronectin increases VEGF-C expression in colorectal carcinoma involving the PI3K/AKT signaling pathway. *PloS One*. 2012;7(4):e35378.
36. Yousif NG. Fibronectin promotes migration and invasion of ovarian cancer cells through up-regulation of FAK-PI 3 K/A kt pathway. *Cell Biology International*. 2014;38(1):85-91.
37. Moosakhani A, Gaeini AA, Kordi M, Ravasi A. Examine Eight Weeks of Voluntary Exercise in Enriched Environment on the Expression of PI3K, Akt, Amyloid Beta Accumulation and Cell Death Rate in the Hippocampus of Male Wistar Rats with Alzheimer's Disease. *Journal of Sport Biosciences*. 2024.



38. Zarin Afzal M, Kazemzadeh Y, Sedaghati S, Mirzaiyan S, Banaeifar A. The effect of interval training with curcumin supplementation on LRP1 and beta amyloid levels in rats with Alzheimer's disease. *Ebnesina*. 2021;23(4):4-14.
39. Mohammadi ZF, Ebrahimzadeh M. The Effect of Six Weeks-Voluntary Wheel Running on Brain Amyloid Beta (1-42) Levels of Diabetic Rats. *Zahedan Journal of Research in Medical Sciences*. 2013;15(5).
40. Nabizadeh F. Neuroprotective role of Fisetin in Alzheimer's disease: An overview of potential mechanism and clinical findings. *Neurology Letters*. 2024;3(Special Issue (Diagnostic and Therapeutic advances in Neurodegenerative diseases)):14-25.